

Embryonic and Fetal Adipogenesis

Emilio Herrera*

University San Pablo CEU, Madrid, Spain

***Corresponding Author:** Emilio Herrera, University San Pablo CEU, Madrid, Spain.

Received: August 27, 2019; **Published:** September 05, 2019

Adipogenesis is the process by which undifferentiated precursor cells (preadipocytes) get differentiated into mature adipocytes. Adipose tissue originates in the embryonic mesoderm that contains a variety of cells: mesenchymal cells, preadipocytes, fibroblasts and adipocytes. In human embryo and fetuses adipose tissue progressively develops from the 14th to the 24th weeks of gestation strongly associated with the formation of blood vessels, and fat lobules are the earliest identified structures before typical vacuolated fat cells appear [1]. White adipocytes are the primary cell type in mammals for the energy storage and mobilization in the form of tricylglycerides and have paracrine and endocrine functions through the synthesis and secretion of adipokines and growth factors whereas brown adipocytes have abundance of mitochondria and actively contribute to energy expenditure by the expression of the mitochondrial uncoupling protein [2]. It was generally assumed that brown and white adipocytes were derived from a common adipogenic precursor, although by using a primary cell model from mice it was concluded that brown preadipocytes have a distinct origin that white adipose tissue [3]. However, although the process of adipogenesis is still minimally characterized, by studying human fetal mesenchymal stem cells differentiation it has been concluded that brown adipocytes are indeed derived from the mesenchymal lineage, along with white adipocytes [4].

Adipogenesis is a tightly regulated cellular differentiation process that requires the sequential activation of several transcription factors. The process of adipogenesis requires highly organized and controlled expression of a cascade of transcription factors in the preadipocytes, which are regulated by hormones, nutrients and epigenetic factors. Embryonic and fetal adipose tissue precursors and regulatory mechanisms appear distinct from adults. The principal regulator of adipocyte differentiation is the nuclear hormone receptor PPAR γ , which induction and maintenance in adults is facilitated by other transcription factor such as C/EBP α [5], which is the founding member of the CCAAT enhancer-binding protein family. The coordination of PPAR γ with C/EBP α maintains adipocyte gene expression in adults [6]. However, in mice it has been shown that the precursor cells giving rise to mature adipocytes during embryogenesis is fully C/EBP α independent [7]. As recently reviewed [8], from studies in mice, embryonic and fetal adipogenesis requires Zfp423 which is a multi zinc-finger transcription factor that is expressed in preadipocytes and mature adipocytes and has been found to be essential for terminal differentiation of adipocytes during fetal adipose tissue development [9]. These transcription factors participate in a single pathway of fat cell development with PPAR γ being the proximal effector of adipogenesis [10]. The activity of PPAR γ is modulated by a gene regulation cascade of selected corepressors and coactivators like steroid receptor coactivator 1 (SRC1), nuclear receptor corepressor (NCoR), the silencing mediator for retinoid and thyroid hormone receptor (SMRT) and an NAD⁺-dependent histone deacetylase and chromatin-silencing factor (SIRT1) which have been proposed to control adipogenesis [11,12].

By studying transgenic mice it has been shown that embryonic preadipocytes that proliferate during the prenatal period expressed perilipin and adiponectin [13], which is at odds with the previous believe that these markers are exclusively expressed in fully differentiated adipocytes in adults [14,15].

Finally, it must be recognized that the origin and development of adipose tissues during intrauterine life is still poorly understood. Moreover, although most of what we know about adipogenesis comes from studies of rodents either in vivo or in cell culture there are examples of adipose tissue function that do not translate from rodents to the human. Therefore, careful interpretation of the findings from experimental animal models should be taken into account for their extrapolation to the human fetal adipogenesis. During pregnancy, maternal health conditions and nutrition state are known to influence the developmental patterning of adipose tissue with lifelong impli-

cations and given the endocrine function of adipose tissue and that alterations of adipocyte development promote metabolic disease state in adults future studies in the area of embryonic and fetal adipogenesis is warranted.

Bibliography

1. Poissonnet CM., *et al.* "The chronology of adipose tissue appearance and distribution in the human fetus". *Early Human Development* 10.1-2 (1984): 1-11.
2. Cannon B and Nedergaard J. "Brown adipose tissue: function and physiological significance". *Physiological Reviews* 84.1 (2004): 277-359.
3. Timmons JA., *et al.* "Myogenic gene expression signature establishes that brown and white adipocytes originate from distinct cell lineages". *Proceedings of the National Academy of Sciences of the United States of America* 104.11 (2007): 4401-4406.
4. Morganstein DL., *et al.* "Human fetal mesenchymal stem cells differentiate into brown and white adipocytes: a role for ERRalpha in human UCP1 expression". *Cell Research* 20 (2010): 434-444.
5. Hu E., *et al.* "Transdifferentiation of myoblasts by the adipogenic transcription factors PPAR gamma and C/EBP alpha". *Proceedings of the National Academy of Sciences of the United States of America* 92.21 (1995): 9856-9860.
6. Cristancho AG and Lazar MA. "Forming functional fat: a growing understanding of adipocyte differentiation". *Nature Reviews Molecular Cell Biology* 12.11 (2011): 722-734.
7. Wang QA., *et al.* "Distinct regulatory mechanisms governing embryonic versus adult adipocyte maturation". *Nature Cell Biology* 17.9 (2015): 1099-1111.
8. Ghaben AL and Scherer PE. "Adipogenesis and metabolic health". *Nature Reviews Molecular Cell Biology* 20.4 (2019): 242-258.
9. Shao M., *et al.* "Fetal development of subcutaneous white adipose tissue is dependent on Zfp423". *Molecular Metabolism* 6.1 (2017): 111-124.
10. Rosen ED., *et al.* "C/EBPalpha induces adipogenesis through PPARgamma: a unified pathway". *Genes and Development* 16.1 (2002): 22-26.
11. Farmer SR. "Transcriptional control of adipocyte formation". *Cell Metabolism* 4.4 (2006): 263-273.
12. Kim TH., *et al.* "Modulation of the transcriptional activity of peroxisome proliferator-activated receptor gamma by protein-protein interactions and post-translational modifications". *Yonsei Medical Journal* 54.3 (2013): 545-559.
13. Hong KY., *et al.* "Perilipin+ embryonic preadipocytes actively proliferate along growing vasculatures for adipose expansion". *Development* 14.25 (2015): 2623-2632.
14. Greenberg AS., *et al.* "Perilipin, a major hormonally regulated adipocyte-specific phosphoprotein associated with the periphery of lipid storage droplets". *Journal of Biological Chemistry* 266.17 (1991): 11341-11346.
15. Cawthorn WP., *et al.* "Adipose tissue stem cells meet preadipocyte commitment: going back to the future". *Journal of Lipid Research* 53.2 (2012): 227-246.

Volume 4 Issue 8 October 2019

© All rights reserved by Emilio Herrera.